

A Review of Bosnia and Herzegovina Medicinal Plants with Promising Antiviral Properties

Tutmić, L.¹, Parić, A.¹, Jerković-Mujkić, A.¹, Muratović, E.^{1}*

¹ University of Sarajevo - Faculty of Science, Department of Biology, Laboratory for Research and Protection of Endemic Resources, Zmaja od Bosne 33-35, 71000 Sarajevo, Bosnia and Herzegovina
[*edina.muratovic@pmf.unsa.ba](mailto:edina.muratovic@pmf.unsa.ba)

Abstract

The study explored the antiviral potential of ethnobotanical plants in Bosnia and Herzegovina, aiming to identify plants with promising antiviral properties and commonly used plant organs and preparations. Three plant families, Lamiaceae, Compositae and Rosaceae showed significant antiviral potential based on the number of species mentioned in the literature. The most studied viruses were herpes simplex virus (HSV), influenza virus (Influenza), and SARS-CoV-2. Tea was the most popular herbal preparation, and aerial parts and leaves were the most frequently used for therapeutic applications. These findings highlight the potential of plants as natural sources of antiviral compounds, paving the way for further research and new therapeutic approaches.

Keywords: ethnobotany, plants, antiviral potential, viruses, plant parts, herbal preparations, Bosnia and Herzegovina.

Received: 21 October 2024; revision received: 30 October 2024; accepted: 1 November 2024. Editor in Chief: prof. dr. Samir Đug.

1. Introduction

Viral infections present a significant health problem due to the high rate of morbidity and mortality. Numerous viruses, such as human immunodeficiency virus (HIV), hepatitis A, B and C (HAV, HBV and HCV), and influenza virus, have been virulent and life-threatening pathogens for decades.

Viral infections have caused many pandemics throughout world history. The deadliest pandemic, the Spanish flu of 1918, infected around 500 million people worldwide and have killed 20 to 50 million. Other notable pandemics include the Ebola virus, between 2013 and 2016, resulting in 11,323 deaths (Trilla et al., 2008), as well as the Severe Acute Respiratory Syndrome (SARS) in 2003, the Middle East Respiratory Syndrome (MERS) in 2015 (Zumla et al., 2015), and in 2019 Coronavirus (COVID-19) pandemic, which has led to more than 776,471,644 infections and over 7,068,677 deaths (World Health Organization, 2024).

The major portion of diseases that do not have a cure are viral diseases (Choudhary et al., 2024). In recent decades, advanced scientific research has led to the discovery of many synthetic antiviral agents that effectively combat various viral infectious diseases. These synthetic agents have been reported to cause numerous adverse effects and become ineffective against new virus strains over time (Kurokawa et al., 2010). Even though more and more antiviral drugs have been approved in clinic studies, long-term use can easily lead to the emergence of drug resistance and side effects (Zhao et al., 2023). One of the global problems of viral infections is the cost of treatment, and there is a need to develop new strategies that will enable finding affordable and effective antiviral drugs (Bachar et al., 2021). The emergence of the severe acute respiratory syndrome-coronavirus (SARS-CoV)-2 pandemic has highlighted our ability to swiftly and resolutely respond to a potential outbreak (Tournier and Kononchik, 2021). As a result, the search for new antiviral agents is now focusing not only on synthetic combinations but also on plant-origin metabolites (Perera and Efferth, 2012).

Medicinal plants have been used for centuries for healing, although their use has not always been scientifically supported. Today, they form the basis of the system of traditional medicine. Traditional medicine includes health practices, approaches, knowledge and beliefs about herbal, animal and mineral drugs, spiritual treatments, and manual procedures applied individually or in combination to diagnose, treat or prevent disease or maintain health (World Health Assembly, 2003). This knowledge is consolidated and available to the public thanks to ethnobotanical research. In recent years, scientists, along with new technologies and research methods, have been discovering new bioactive components and therapeutic possibilities of plants,

contributing to the development of innovative solutions in the treatment and prevention of diseases. This constant progress in plant research enables the wider use of herbal products in modern medicine (Beressa et al., 2021, Najmi et al., 2022).

This study aimed to summarize data from the literature on the antiviral effects of plants that were/are traditionally used in Bosnia and Herzegovina, as well as investigate the relationship between the traditional use of these plants and their potential antiviral properties.

2. Material and Methods

In this retrospective analysis, previously published ethnobotanical studies were used to gain insight into the use of plants in traditional medicine in Bosnia and Herzegovina (Zovko, 1890; Glück, 1892; Fazlagić, 1894, 1895; Kulinović, 1898; Bratić, 1903, 1908; Medić, 1904 a, b; Dragičević, 1909; Filipović-Fabijanić, 1964; Filipović-Fabijanić, 1968; Filipović-Fabijanić, 1969/1970, 1971; Redžić, 2007, 2010; Šarić-Kundalić et al., 2010 a,b, 2011, 2015, 2016; Fereier et al., 2015; Ginko et al., 2023); as well as the study Muratović and Parić (2023) combined these studies to provide an overview of plant species used throughout history until today. Relevant databases, like Web of Science, Scopus, and ScienceDirect (Elsevier), were used to search for medicinal plants with antiviral activity using keywords such as "antiviral activity" and "bioactive compounds."

3. Results

Out of the total number of 145 ethnobotanically researched and documented plant species (Muratović and Parić, 2023), this paper focuses on 56 taxa scientifically proven to have antiviral potential in addition to ethnobotanical use (Table 1.). This selection provides a better understanding of plants that, based on traditional knowledge, were used to treat various diseases and showed significant potential in combating viral infections.

Based on the data in (Table 1.), research has been conducted on the antiviral properties of various plant families, and some of them were dominant. The most prominent family was Lamiaceae, with ten species investigated for their antiviral potential. The Asteraceae family takes second place with eight plant species, while Rosaceae ranks third with six species. The Fabaceae family follows them with three, and the rest of the families have two or one plant species each.

Table 1. Medicinal plants in traditional use in Bosnia and Herzegovina (Muratović and Parić, 2023) and their antiviral potential

Scientific Name		Family	Vernacular name	Part(s) of utilization	Therapeutic uses	Mode of utilization	Antiviral potential	References
<i>Agrimonia eupatoria</i> L.		Rosaceae	petrovac, turica, turika	aerial part	anti-haemorrhagic, anti-inflammatory, diarrhea, hepatics, rheumatism	infusion		
<i>Alchemilla vulgaris</i>		Rosaceae	vrkuta, gospin plašt, plahtica	aerial part	anti-hemorrhagic, anti-inflammatory, antiseptic, dysmenorrhea, hormonal disorders	infusion	HBV*	Kwon et al., 2005
<i>Angelica archangelica</i> L.	Compositae	Apiaceae	andelika	aerial part	analgesic, asthma, flatulence, migraine	infusion		
<i>Anthyllis vulneraria</i> L.		Fabaceae	ranjenik	aerial part	contusion, cough, skin disorders, wound	infusion		
<i>Arctium lappa</i> L.		Compositae	čičak, repuh	root, leaf, fruit	antipruritic, aperitif, arteriosclerosis, diabetes, diuretic, eczema, hepatic, hyperglycemia, psoriasis	infusion		
<i>Artemisia vulgaris</i> L.			divlji pelin, komonjika, komunika, crnobilj, crni pelin, umit	young shoots	antianemic, antipyretic, antispasmodic, aperitif, diabetes, diarrhea, diuretic, gastritis, hepatic, laxative, rheumatism	infusion, tincture	HSV-1 EVC HSV IAV	HSV-1 CVB3 SARS-CoV-2 Rajtar et al., 2017 Suručić et al., 2022
Xiao, 2023	Dias et al., 2017			Suganda et al., 1983				

Scientific Name	Family	Vernacular name	Part(s) of utilization	Therapeutic uses	Mode of utilization	Antiviral potential	References
<i>Glycyrrhiza glabra</i> L.	Fabaceae	sladić	root	hepatitis, hysteria, stomach disorders, sore throat, stone and sand in the kidney.	infusion, extract in oil olive	HSV HIV HBV SARS-CoV-2	Sato et al., 1996; Gotoh et al., 1987; Takahara et al., 1994;
<i>Hedera helix</i> L.	Araliaceae	brišjan	leaf	anti-emetic, anti-inflammatory, diuretic, hypertension, rheumatism.	infusion	IAV	
<i>Helichrysum italicum</i> (Roth) G. Don	Compositae	smilje	flower	antianemic, antipyretic, migraine.			
<i>Hypericum perforatum</i> L.	Clusiaceae	kantarion, bogorodičina trava, ivanova trava	aerial part	analeptic, antianemic, antiinfective, antifungal, asthma, diarrhea, dysmenorrhea, gastritis, hepatic, migraine, rheumatism.	infusion		
<i>Juglans regia</i> L.	Juglandaceae	orah	leaf, immature fruits	dermatitis, diabetes, gastritis, hypo-hyperthyroidism, rheumatism.	infusion, syrup, honey	HIV	
				HIV EV/C CVB1 HAdV	SARS-CoV-2		
				Mohamed et al., 2022; Zhang et al., 2017	Chien et al., 2004	Hong et al., 2015	D'Angeli et al., 2021

Scientific Name		Family	Vernacular name	Part(s) of utilization	Therapeutic uses	Mode of utilization	Antiviral potential	References
<i>Juniperus communis</i> L.		Cupressaceae	kleka, smreka	galbulus, yang shots	antianemic, antiinfective, anti-inflammatory, diabetes, dyspepsia, gastritis, hepatics, laxative, rheumatism, scurvy.			
<i>Laurus nobilis</i> L.		Lauraceae	lovor	leaf, fruit	antitussive, flatulence, gastritis.		IAV	
<i>Lavandula angustifolia</i> Mill. subsp. <i>angustifolia</i>		Lamiaceae	despik, lavanda	aerial part	analeptic, cardiotonic, gastritis, hypertension, migraine, rheumatism	infusion		
<i>Marrubium vulgare</i> L.	Compositae	Lamiaceae	obična marulja, očajnica, smrduša	aerial part	antiseptic	infusion, oil		
<i>Matricaria chamomilla</i> L.		kamilica	leaf, flower		antianemic, anti-inflammatory, antipyretic, arthritis, antispasmodic, dermatitis, diarrhea, diuretic, flatulence, gastritis, hypertension, migraine.	HSV1 SARS-CoV-2	IAV	Najar et al., 2022
				infusion				Suganda et al., 1983
				HSV EVC				Fayyad et al., 2014
					Abou et al., 2021			Khodja et al., 2023

Scientific Name	Family	Vernacular name	Part(s) of utilization	Therapeutic uses	Mode of utilization	Antiviral potential	References
<i>Melissa officinalis</i> L.	Lamiaceae	melisa, matičnjak, pčelinja ljubica, limunčić	aerial part	analeptic, antiviral, asthma, cardiotonic, dyspepsia, gastritis, hypertension, laxative.			
<i>Mentha pulegium</i> L.	Lamiaceae	verem trava	aerial part	analeptic, anti-inflammatory, gastritis.			
<i>Nigella damascena</i> L.	Ranunculaceae	čurokot, mačkov brk	seed	analeptic, hepatic.			
<i>Origanum vulgare</i> L.	Lamiaceae	divlji origano, mravinač, vranilova	aerial part	analeptic, antipyretic, dyspepsia, gastritis, hepatic.			
<i>Petasites hybridus</i> (L.) G. Gaertn. & al.	Compositae	lopuh, veliki čičak, repuh	root, leaf	rheumatism	infusion	HSV HIV SARS-CoV-2	
<i>Plantago media</i>	Plantaginaceae	trputac, srednja bokvica	leaf	antianemic, anti-hemorrhagic, anti-inflammatory, antipyretic, diarrhea, gastritis, hepatics, rheumatism.	infusion		
Zhakipbekov et al., 2023; Musarra-Pizzo et al., 2021	HSV HAdV	SARS-CoV-2	SARS-CoV-2	HSV			Behzadi et al., 2023
	Urda et al., 2022	Zhang et al., 2014	Imran et al., 2022	Parsania et al., 2017			

Scientific Name		Family	Vernacular name	Part(s) of utilization	Therapeutic uses	Mode of utilization	Antiviral potential	References
<i>Polygonatum odoratum</i>		Asparagaceae	pokosnica	rhizome	bleeding from trachea, cough, hemorrhoids, leucorrhea, liver disorders, uterus disorders, wound.	fresh juice, infusion	IAV	Pang et al., 2020
<i>Primula veris</i>		Primulaceae	jaglac, jaglika	aerial part	analeptic, migraine, rheumatism.	infusion	IAV	
<i>Prunella laciniata</i>		Lamiaceae	bijela celinčica	aerial part	asthma, diarrhea, liver, strengthen of heart	decoction, infusion		
<i>Punica granatum</i>		Lythraceae	nar	bark, fruit	diarrhea			
<i>Rosmarinus officinalis</i>		Lamiaceae	ruzmarin, zimrad	leaf	analeptic, cardiotonic, gastritis, hepatics, rheumatism	infusion		
<i>Rubus idaeus</i>		Rosaceae	malina	leaf	antiinfective, gastritis	infusion		
<i>Ruta graveolens</i>		Rutaceae	ruta	aerial part	cardiotonic, laxative, rheumatism	infusion	IAV	
<i>Salvia officinalis</i>	Lamiaceae		kadulja, žalfija	aerial part	anti-inflammatory, antipyretic, diabetes	IAV	Abou Baker et al., 2021	Nikolaeva-Glomb et al., 2013; Moradi et al., 2011; Arunkuma et al., 2018; Oh et al., 2011; Eliopoulos et al., 2022

Scientific Name		Family	Vernacular name	Part(s) of utilization	Therapeutic uses	Mode of utilization	Antiviral potential	References
<i>Sambucus ebulus</i>		Viburnaceae	aptovina, apt, burjan	leaf, root, fruit	antipruritic			
<i>Sinapis arvensis</i>		Brassicaceae	gorušica, hardala	seed	asthma			
<i>Taraxacum</i> F. H. Wigg. sect. <i>Taraxacum</i>		Compositae	maslačak, žućanik	root, leaf, flower	antianemic, antispasmodic, diabetes, eczema, hemorrhoids, hepatic, hepatitis, laxative, rheumatism			
<i>Teucrium chamaedrys</i>	Lamiaceae	dupčać, stupčać, mravak, suhovrh	aerial part	antibacterial, diabetes, diarrhea, gastritis		infusion, honey	juice	
<i>Urtica dioica</i>	Urticaceae	žara, kopriva	whole plant	analgesic, antianemic, antipyretic, arteriosclerosis, dermatitis, diabetes, diuretic, dysmenorrhea, hepatics, rheumatism	infusion	HCV	HSV1	
					HIV			
					Todorov et al., 2015	Sharifi-Rad et al., 2017	Ghaffari et al., 2021	
Uncini et al., 2005								

Scientific Name		Family	Vernacular name	Part(s) of utilization	Therapeutic uses	Mode of utilization	Antiviral potential	References
<i>Vaccinium myrtillus</i>		Ericaceae	borovnica, vrisinje	leaf, fruit	antianemic, anti-inflammatory, arteriosclerosis, diabetes, diarrhea, dyspepsia, gastritis, vitamin C and E deficiencies	infusion, juice	IAV CVB1 EVC SARS-CoV-2	Nikolaeva-Glomb et al., 2013
<i>Vaccinium vitis-idaea</i>		Ericaceae	brusnica	leaf, fruit	antiinfective, diuretic, rheumatism	infusion	IAV	
<i>Veronica officinalis</i>		Plantaginaceae	čestoslavica, dobričica	leaf, flower	antipruritic, gastritis, rheumatism	infusion		
<i>Viola odorata</i>	Violaceae	Violaceae	ljubica mirisna	leaf, flower	antitussive			
<i>Viscum album</i>	Loranthaceae	imela bijela	aerial part	analgesic, cardiotonic, dysmenorrhea, hypertension	infusion, tincture	SARS-CoV-2	IAV	
<i>Vitex agnus-castus</i>	Lamiaceae	fratarski biber, konopljika	aerial part, seed	against high potency, for nervous balance, hysteria, menstrual disorders	infusion, ecocin, powder	HSV HIV	HPIV	Adel Mehraban et al., 2023
		Karagöz et al., 2003				Mazurkova et al., 2020		Islam et al., 2024

*Acronym of virus: HBV (*Hepatitis B virus*), HCV (*Hepatitis C virus*), SARS-CoV-2 (*Severe Acute Respiratory Syndrome Coronavirus 2*), HSV-1 and 2 (*Herpes simplex virus 1 and 2*), EV (*Enterovirus*), CVB1 and B3 (*Coxsackie virus B1 and B3*), EVC (*poliovirus-Enterovirus C*), IAV (*Influenza A virus*), HIV (*Human immunodeficiency virus*), HPV 1 (*Human papilloma virus*), HPV (*Human papilloma virus 2*), HAdV (*Human adenovirus*), HPIV (*Human parainfluenza virus*)

The most common form of utilization (Table 1.) is infusion, for which 53 plant species are used. Infusion is prepared by pouring hot water over plant parts, and it is one of the simplest and most widespread ways of preparing medicinal plants. Nine plant species are used for fresh juice. It is frequently prepared from fresh parts of the plant, such as leaves, stems or roots, and is used due to the high concentration of bioactive substances. The tincture, which involves alcohol to extract the plant components, is prepared from four plant species. Three plant species are used for decoction (the plant material boiled in water), two for syrup, honey and oil, and one for honey. These data clearly showed that infusion dominates, while the other forms have less frequent usage.

The distribution of bioactive compounds varies in different parts of plants. According to (Table 1.), it was obvious that in folk medicine, the aerial part of plants is predominantly utilized (confirmed for 22 plant species). Additionally, leaves of 21 species are used in preparations, fruits of 10, and flowers and roots of seven species. Less commonly used plant organs, such as tree bark and rhizomes, were found to be employed for three taxa and young shoots for two. Finally, unripe fruits, seeds, fresh juice, tree sap and the whole plant were confirmed for only one plant species.

Based on the available literature data on the antiviral potential of plants (Table 1.), we selected the most relevant viruses against which these plants showed activity. Among the findings, 28 plant species showed antiviral potential against the herpes simplex virus (HSV-1 and HSV-2), 17 showed an inhibitory effect on the influenza virus, 13 on SARS-CoV-2, and 11 against the human immunodeficiency virus (HIV), etc.

In the reviewing literature, the Lamiaceae family showed the most pronounced antiviral potential on 13 different viruses, followed by Compositae, Rosaceae, Fabaceae, etc. (Table 1.).

4. Discussion

Regarding the antiviral properties of plant families, Lamiaceae, Compositae, and Rosaceae are the most extensively studied. It was expected since they are rich in taxa and contain various bioactive compounds known for their antiviral properties. The leaves of these families' members are sources of flavonoids and phenolic acids, such as 4-Hydroxybenzoic acid, vanillic acid, and chlorogenic acid (Sytar et al., 2018).

The plants from these families have been used in folk medicine for centuries, and modern research has confirmed their effectiveness in combating viruses. Behzadi et al. (2023) reported that *Melissa officinalis* has an antiviral effect against HSV, HIV,

SARS-CoV-2, CPV, HBV, and HCV (Table 1.). Additionally, its effectiveness as an alternative therapy for several viral infections has been confirmed by Muratović and Parić (2023). *M. officinalis* is rich in organic active compounds, mainly essential oils (including citronellal, menthol, eugenol, linalool, and geraniol) and polyphenols (including rosmarinic acid, caffeic acid, chlorogenic acid, salicylic acid, ellagic acid, and quercetin). Its significant antiviral effect is due to the presence of rosmarinic acid in its methanol extract (Chen et al., 2017).

Another example is *Vaccinium myrtillus*, a species with significant antiviral potential against various viruses such as poliovirus type 1 (PV-1), coxsackievirus B1 (CV-B1), human respiratory syncytial virus A2 (HRSV-A2), influenza virus A/H3N2, SARS-CoV-2, and Coxsackie virus (Nikolaeva-Glomb et al., 2014). This species contains phenolic compounds, including chlorogenic acid, quercetin and arbutin, which have outstanding antioxidant capacity both in vitro and in vivo. These compounds are present in different representatives of the Ericaceae family (Colak et al., 2017; Štefănescu et al., 2019). The *Juglans regia* has a wide range of applications in daily life (Muratović and Parić, 2023). This taxon has been proven to have antiviral activity against HIV, Poliovirus, Coxsackievirus B1, Adenovirus, Sindbis virus (SINV), and herpes simplex virus (HSV) due to its active components quercetin, myricetin, and naringenin (Bhat et al., 2023). Additionally, *Glycyrrhiza glabra* has been found to have significant antiviral effects against HSV, HIV, HBV, SARS2, and H5N1 viruses due to its active compounds glycyrrhizin and glycyrrhetic acid, which belong to the group of triterpenoids (Wang et al., 2015).

Although more than 200 human viruses have been discovered, antiviral medications are available only for a limited number of infections that target proteins encoded by a single virus (such as influenza A and B, hepatitis B and C, herpes, and HIV), providing a narrow spectrum of coverage (Karim et al., 2023). The emergence or re-emergence of viruses with epidemic or pandemic potential, such as Ebola, SAR-CoV-1, and SAR-CoV-2, has repeatedly reminded us of the importance of developing broad-spectrum antiviral drugs (Geraghty et al., 2021). In this context, our results highlight the diversity and complexity of antiviral potential among different plant families.

Table 2 shows the relationship between ethnobotanical research in Bosnia and Herzegovina and the research about the antiviral potential of selected plants. Local communities noticed that certain plant species help in the treatment of specific symptoms. Microbiological studies have proven that the plants used in traditional medicine also have antiviral potential and that the symptoms caused by these viruses can be identified with the symptoms recognized by folk medicine for the same plant species.

One of the examples is the already mentioned blueberry, which has proven antiviral potential against the SARS-CoV-2 virus, whose most common symptoms are hemorrhage, high temperature, diarrhea, anemia, sore throat, cough and diabetes (Table 2.). At the same time, it is important to emphasize that in the traditional medicine of B&H, this plant is used for antianemic, anti-inflammatory, arteriosclerosis, diabetes, diarrhea, dyspepsia, gastritis, vitamin C and E deficiency (Table 1.).

However, it is important to emphasize that the symptoms of viral infection are often subjective and can vary from person to person. For example, hepatitis B virus and hepatitis C virus can cause chronic infections. Viral hepatitis can cause a wide spectrum of clinical presentations from a benign form with quite mild or no symptoms to acute liver failure and sometimes liver cancer or death (Cheung and Kwo, 2020).

Various host factors collectively shape an individual's response to pathogens such as age, genetic factors, immune system function, nutritional status, microbiome, underlying health conditions as well as environmental factors including living conditions, socioeconomic status and exposure to pollutants and various other aspects (Hayward, 2023). Therefore, these results highlight the complexity of the relationship between traditional knowledge and modern scientific findings in the context of the treatment of viral infections.

Table 2. Antiviral potential of the plants with ethnobotanical use in Bosnia and Herzegovina

Virus (symptoms)	Plant species (Muratović et Parić 2023)	Reference
Herpes simplex virus (skin disorders, eczema, dermatitis, sore throat, inflammation, elevated temperature, migraine, asthma, diarrhea, gastritis)	<i>Angelica archangelica, Anthyllis vulneraria, Arctium lappa, Artemisia vulgaris, Betula pendula, Ceratonia siliqua L, Chelidonium majus, Cornus mas, Fagus sylvatica, Ficus carica, Foeniculum vulgare, Glycyrrhiza glabra, Laurus nobilis, Melissa officinalis, Matricaria chamomilla, Mentha pulegium, Plantago media, Punica granatum, Sinapis arvensis, Teucrium chamaedrys</i>	Xiao et al., 2024; Banerjee et al., 2007; McMillan et al., 1993; Napier et al., 2018; Zhang et al., 2022; Colemont et al., 1990

Influenza virus (high fever, anemia, diarrhea, cough)	<i>Arctium lappa, Foeniculum vulgare, Fragaria vesca, Hedera helix, Juniperus communis, Polygonatum odoratum</i> (Mill.), <i>Punica granatum, Rubus idaeus, Salvia officinalis, Vaccinium myrtillus, Vaccinium vitis-idaea, Veronica officinalis</i>	InformedHealth, 2006; Rice et al., 1998; Boktor et al., 2024
Severe acute respiratory syndrome coronavirus 2 (hemorrhages, high fever, diarrhea, anemia, sore throat, cough, diabetes)	<i>Alchemilla vulgaris, Foeniculum vulgare, Glycyrrhiza glabra, Hypericum perforatum, Laurus nobilis, Vaccinium myrtillus, Viola odorata</i>	Steenblock et al., 2023; Cornea et al., 2022; Bergamaschi et al., 2021; Juthi 2023; Waliszewska-Prostó et al., 2021; Rai et al., 2023
Human immunodeficiency virus (hypertension, diarrhea, dermatitis, fever and elevated temperature)	<i>Betula pendula, Chelidonium majus, Cornus mas, Filipendula vulgaris, Helichrysum italicum, Hypericum perforatum, Juglans regia, Prunella laciniata, Urtica dioica</i>	Nguyen et al., 2022; Garg and Snke, 2017; Harimenshi et al., 2022;
Poliovirus (flu-like symptoms, cough)	<i>Fragaria vesca, Matricaria chamomilla, Vaccinium myrtillus</i>	Wolbert et al., 2024
Coxsackie virus (dermatitis, diarrhea)	<i>Juglans regia, Vaccinium myrtillus</i>	Valestra et al., 2016; Trayer and Gore, 2020
Hepatitis B virus (hepatitis, stomach problems)	<i>Agrimonia eupatoria, Glycyrrhiza glabra</i>	Liu et al., 2022
Human papillomavirus (no data)	No data	No data
Adenovirus (gastrointestinal disturbances)	<i>Juglans regia, Plantago media</i>	Kumthip et al., 2019
Enterovirus (No data)	No data	No data
Hepatitis C virus (hepatitis)	<i>Taraxacum</i> sp.	Basit et al., 2023
Human parainfluenza virus (no data)	No data	No data

Among the plants that have been used throughout history in folk medicine and today are completely forgotten, there are the following: *Anagallis arvensis*, *Fritillaria meleagris*, *Galanthus nivalis*, *Hioscyamys niger*, *Rubia tinctorum* (Muratović and Parić, 2023). The antiviral effect of these plants was proven in different studies. Thus, the *Anagallis arvensis* has an antiviral potential against Herpes simplex and Poliovirus (Amoros et al., 1987), *Fritillaria meleagris* against the Influenza virus (Kim et al., 2020), *Galanthus nivalis* against Hepatitis C virus (Ashfaq et al., 2011), *Hioscyamys niger* against Respiratory Syndrome Coronavirus 2 (Kosari et al., 2021, 2024) and *Rubia tinctorum* against rotavirus (Sun et al., 2016).

5. Conclusions

In recent years, there has been a growing interest and need for antiviral drugs due to the latest pandemic. Consequently, medicinal plants are receiving increased attention. Research has demonstrated that the long-standing use of medicinal plants in folk medicine can offer valuable insights for further studies into their active

components. This study highlights the importance of a multidisciplinary approach, which can provide a broader perspective and better solutions for this and other issues.

6. References

1. Abe, N., Ebina, T., & Ishida, N. (1982). Interferon induction by glycyrrhizin and glycyrrhetic acid in mice. *Microbiology and Immunology*, 26, 535–539. doi: 10.1111/j.1348-0421.1982.tb00207.x.
2. Abou Baker, D.H., Amarowicz, R., Kandeil, A., Ali, M.A., & Ibrahim, E.A. (2021). Antiviral activity of *Lavandula angustifolia* L. and *Salvia officinalis* L. essential oils against avian influenza H5N1 virus. *Journal of Agriculture and Food Research*, 4:100135. doi: 10.1016/j.jafr.2021.100135.
3. Adel Mehraban, M.S., Shirzad, M., Mohammad Taghizadeh Kashani L., Ahmadian-Attari M.M., Safari A.A., Ansari N., Hatami H., & Kamalinejad M. (2023). Efficacy and safety of add-on *Viola odorata* L. in the treatment of COVID-19: A randomized double-blind controlled trial. *Journal of Ethnopharmacology*, 25;304:116058. doi: 10.1016/j.jep.2022.116058.
4. Aiken, C., & Chen, C.H. (2005). Betulinic acid derivatives as HIV- 1 antivirals, *Trends in Molecular Medicine*, 11(1)31-36. doi: 10.1016/j.molmed.2004.11.001.
5. Alhadrami, H.A., Sayed, A.M., Hassan, H.M., Youssif, K.A., Gaber, Y., Moatasim, Y., Kutkat, O., Mostafa, A., Ali, M.A., Rateb, M.E., Abdelmohsen, U.R., & Gamaleldin, N.M. (2021). Cnicin as an Anti-SARS-CoV-2: An Integrated In Silico and In Vitro Approach for the Rapid Identification of Potential COVID-19 Therapeutics. *Antibiotics*, 7;10(5):542. doi: 10.3390/antibiotics10050542.
6. Al-Megrin, W.A., AlSadhan, N.A., Metwally, D.M., Al-Talhi, R.A., El-Khadragy, M.F., & Abdel-Hafez, I.J.M. (2020). Potential antiviral agents of *Rosmarinus officinalis* extract against herpes viruses 1 and 2. *Bioscience Reports*, 26;40(6):BSR20200992. doi: 10.1042/BSR20200992.
7. Amoros, M., Fauconnier, B., & Girre, R.L. (1987). In vitro antiviral activity of a saponin from *Anagallis arvensis*, Primulaceae, against herpes simplex virus and poliovirus. *Antiviral Research*, 8(1):13-25. doi: 10.1016/0166-3542(87)90084-2.
8. Arunkumar, J., & Rajarajan, S. (2018). Study on antiviral activities, drug-likeness and molecular docking of bioactive compounds of *Punica granatum* L. to Herpes simplex virus - 2 (HSV-2). *Microbial Pathogenesis*, 118:301-309. doi: 10.1016/j.micpath.2018.03.052.
9. Ashfaq, U.A., Masoud, M.S., Khalid, S., Nawaz, Z., & Riazuddin, S. (2011). Inhibition of hepatitis C virus 3a genotype entry through *Glanthus Nivalis* Agglutinin. *Virology Journal*, 20(8):248. doi: 10.1186/1743-422X-8-248.

10. Ay, E., & Duran, N. (2018). Investigation of the antiviral activity of *Ficus carica* L latex against HSV-2.33-37. doi: 10.24264/icams-2018.I.3.
11. Bachar, S.C., Mazumder, K., Bachar, R., Aktar, A., & Al Mahtab, M. (2021). A Review of Medicinal Plants with Antiviral Activity Available in Bangladesh and Mechanistic Insight Into Their Bioactive Metabolites on SARS-CoV-2, HIV and HBV. *Frontiers in Pharmacology*, 12,732891. doi: 10.3389/fphar.2021.732891.
12. Behzadi, A., Imani, S., Deravi, N., Mohammad Taheri, Z., Mohammadian, F., Moraveji, Z., Shavysi, S., Mostafaloo, M., Soleimani Hadidi, F., Nanbakhsh, S., Olangian-Tehrani, S., Marabi, M.H., Behshood, P., Poudineh, M., Kheirandish, A., Keylani, K., & Behfarnia, P. (2023). Antiviral Potential of *Melissa officinalis* L.: A Literature Review. *Nutrition and Metabolic Insights*, 12(16),11786388221146683. doi: 10.1177/11786388221146683.
13. Beressa, T.B., Deyno, S., Mtewa, A.G., Aidah, N., Tuyiringire, N., Lukubye, B., Weisheit, A., Tolo, C.U., & Ogwang, P.E. (2021). Potential Benefits of Antiviral African Medicinal Plants in the Management of Viral Infections: Systematic Review. *Frontiers in Pharmacology*, 24(12),682794. doi: 10.3389/fphar.2021.682794. PMID: 35002686; PMCID: PMC8740180.
14. Bhat, A.A., Shakeel, A., Rafiq, S., Farooq, I., Malik, A.Q., Alghuthami, M.E., Alharthi, S., Qanash, H., & Alharthy, S.A. (2023). *Juglans regia* Linn.: A Natural Repository of Vital Phytochemical and Pharmacological Compounds. *Life (Basel)*, 13(2):380. doi: 10.3390/life13020380.
15. Bratić, T.A. (1903). Pabirci iz narodne medicine u Hercegovini. *Glasnik Zemaljskog Muzeja*. XV/I:153-180.
16. Bratić, T.A. (1908). Narodno liječenje. Iz vremena jedne ljekaruše iz 1843. godine. *Glasnik Zemaljskog Muzeja*. XX/III:343-362.
17. Chen, S.G., Leu, Y.L., Cheng, M.L., Ting, S.C., Liu, C.C., Wang, S.D., Yang, C.H., Hung, C.Y., Sakurai, H., Chen, K.H., & Ho, H.Y. (2017). Anti-enterovirus 71 activities of *Melissa officinalis* extract and its biologically active constituent rosmarinic acid. *Scientific Reports*, 7, 12264. doi: 10.1038/s41598-017-12388-2.
18. Cheung, A., & Kwo, P. (2020). Viral hepatitis other than A, B, and C: evaluation and management. *Clinics in liver disease*, 24(3), 405-419. doi: 10.1016/j.cld.2020.04.008.
19. Chien, N.Q., Hung, N.V., Santarsiero, B.D., Mesecar, A.D., Cuong, N.M., Soejarto, D.D., Pezzuto, J.M., Fong, H.H., & Tan, G.T. (2004). New O-acyl betulinic acids from *Strychnos vanprukii* Craib. *Journal of natural products*, 67, 994–998. doi: 10.1021/np030469i.
20. Choi, J. G., Kim, Y. S., Kim, J. H., & Chung, H. S. (2019). Antiviral activity of ethanol extract of *Geranii Herba* and its components against influenza viruses via

- neuraminidase inhibition. *Scientific reports*, 9(1), 12132. doi: 10.1038/s41598-019-48430-8
21. Choudhary, P., Pant, K., & Kumar, V. (2024). Challenges in development of antiviral drugs and vaccines. In *Pathogenic Viruses and Armamentarium Design* (pp. 157-173). Academic Press.
22. Colak, N., Primetta, A.K., Riihinens, K.R., Jaakola, L., Grúz, J., Strnad, M., Torun, H., & Ayaz, F.A. (2017). Phenolic compounds and antioxidant capacity in different-colored and non-pigmented berries of bilberry (*Vaccinium myrtillus* L.). *Food Bioscience*, 20, 67-78. doi:10.1016/j.fbio.2017.06.004.
23. D'Angeli F., Malfa G.A., Garozzo A., Li Volti G., Genovese C., Stivala A., Nicolosi D., Attanasio F., Bellia F., Ronisvalle S., & Acquaviva R. (2021) Antimicrobial, Antioxidant, and Cytotoxic Activities of *Juglans regia* L. Pellicle Extract. *Antibiotics*, 4;10(2),159. doi: 10.3390/antibiotics10020159
24. Darwish, W.S., Khadr, A.E.S., Kamel, M.A.E.N., Abd Eldaim, M.A., El Sayed, I.E.T., Abdel-Bary, H.M., Ullah, S., & Ghareeb, D.A. (2021). Phytochemical Characterization and Evaluation of Biological Activities of Egyptian Carob Pods (*Ceratonia siliqua* L.) Aqueous Extract: In Vitro Study. *Plants*. 10(12),2626. doi: 10.3390/plants10122626
25. Dias, M.M., Zuza, O., Riani, L.R., de Faria Pinto, P., Pinto, P.L.S., Silva, M.P., de Moraes, J., Ataíde, A.C.Z., de Oliveira Silva, F., Cecílio, A.B., & Da Silva Filho, A.A. (2017). In vitro schistosomicidal and antiviral activities of *Arctium lappa* L. (Asteraceae) against *Schistosoma mansoni* and *Herpes simplex virus-1*. *Biomedicine & Pharmacotherapy*, 94, 489-498. doi: 10.1016/j.biopha.2017.07.116.
26. Dragićević, T. (1909). Narodni lijekovi. *Glasnik Zemaljskog Muzeja*. XXI/III:461-478.
27. Ebrahimi, E., Mousavi-Jazayeri, S.M., Rezaee, M.B., & Parsania, M. (2021). Antiviral Effects of *Aloe vera* (L.) Burm.f. and *Ruta Graveolens* L. Extract on Acyclovir-Resistant Herpes Simplex Virus Type 1. *Journal of Medicinal Plants and By-products*, 1: 103-108. doi: 10.22092/jmpb.2021.352463.1286.
28. Eliopoulos, A.G., Angelis, A., Liakakou, A., & Skaltsounis, L.A. (2022). In Vitro Anti-Influenza Virus Activity of Non-Polar *Primula veris* subsp. *veris* Extract. *Pharmaceuticals*, 15(12), 1513. doi: 10.3390/ph15121513.
29. Fayyad, A.G.S., Ibrahim, N., & Yaakob, W.A. (2014). Phytochemical screening and antiviral activity of *Marrubium vulgare*. *Malaysian Journal of Microbiology*, 10(2), 106-111. doi: 10.21161/mjm.58013.
30. Fazlagić, H. (1894). Narodni lijekovi iz bilinstava u Bosni. *Glasnik Zemaljskog Muzeja*. VI/IV:800-802.

31. Fazlagić, H. (1895). Bajanje. Prilog narodnog lijekarstva Bosne i Hercegovine. Glasnik Zemaljskog Muzeja .VII/I:155-157.
32. Ferrier, J., Saciragic, L., Trakić, S., Chen, ECH., Gendron, R., Cuerrier, A., Balick, M., Redžić, S., Alikadić, E., & Arnason, J. (2015). An ethnobotany of the Lukomir highlanders of Bosnia & Herzegovina. *Journal of Ethnobiology and Ethnomedicine*, 11:81. doi: 10.1186/s13002-015-0068-5.
33. Filipović-Fabijanić, R. (1968). Narodna medicina istočne Hercegovine. *Glasnik Zemaljskog Muzeja N. S. XXIII/Etnologija*: 35-76.
34. Filipović-Fabijanić, R. (1964). Narodna medicina i narodna verovanja (u Žepi). *Glasnik Zemaljskog Muzeja N.S. XIX/Etnologija*:209-236.
35. Filipović-Fabijanić ,R. 1969/1970. O narodnoj medicini stanovništva Lištice s okolinom. *Glasnik Zemaljskog Muzeja N.S. XXIV/XXV/Etnologija*:319-335.
36. Filipović-Fabijanić, R. (1971). "Domaći ljekar" iz 1868. godine sa Širokog Brijega. *Glasnik Zemaljskog Muzeja N.S. XXVI/Etnologija*:117-178.
37. Ghaffari, H., Ataei-Pirkoooh, A., Mirghazanfari, S.M., & Barati, M. (2021). Inhibition of herpes simplex virus type 1 infection by *Sambucus ebulus* extract in vitro. *Medical Journal of The Islamic Republic of Iran*, 35:9, doi: 10.47176/mjiri.35.9.
38. Ghanbari, A., Le Gresley, A., Naughton, D., Kuhnert, N., Sirbu, D., & Ashrafi, G.H. (2019). Biological activities of *Ficus carica* latex for potential therapeutics in Human Papillomavirus (HPV) related cervical cancers. *Scientific Reports*, 9(1):1013. doi: 10.1038/s41598-018-37665-6.
39. Geraghty, R.J., Aliota, M.T., & Bonnac, L.F. (2021). Broad-Spectrum Antiviral Strategies and Nucleoside Analogs. *Viruses*, 13, 667.
40. Ginko, E., Alajmovic Demirović, E., & Šarić-Kundalić, B. (2023). Ethnobotanical study of traditionally used plants in the municipality of Zavidovići, BiH. *Journal of Ethnopharmacology*. 302(Pt A):115888. doi: 10.1016/j.jep.2022.115888.
41. Glück, L. (1892). Narodni lijekovi iz bilinstva u Bosni. *Etnografska študija*. *Glasnik Zemaljskog Muzeja IV/II:134-167*.
42. Gotoh, Y., Tada, K., Yamada, K., Minamitani, M., Negishi, M., Fujimaki, M., Ikematsu, S., Hada, M., Mori, K., Ito, M. & Shigeta, S. (1987). Administration of glycyrrhizin to patients with human immunodeficiency virus infection. *Igaku no Ayumi*, 140, 619-620.
43. Hayward, M. (2023). The Influence of Host Factors on Susceptibility and Resistance to Infectious Diseases. *Journal of Microbiology and Pathology*, 7 (1), 169.
44. Heidary Navid, M., Laszczyk-Lauer, M.N., Reichling, J., & Schnitzler, P. (2024). Pentacyclic triterpenes in birch bark extract inhibit early step of herpes simplex

- virus type 1 replication. *Phytomedicine*, 21;1273-1280. doi: 10.1016/j.phymed.2014.06.007.
45. Hoever, G., Baltina, L., Michaelis, M. Kondratenko, R, Baltina, L, Tolstikov, GA, Doerr, HW, & Cinatl, J. Jr. (2005). Antiviral activity of glycyrrhetic acid derivatives against SARS-coronavirus. *J. Med Chem* 48: 1256–1259. doi: 10.1021/jm0493008.
46. Hong, E.H., Song, J.H., Shim, A., Lee, B.R., Kwon, B.E., Song, H.H., Kim, Y.J., Chang, S.Y., Jeong, H.G., Kim, J.G., Seo, S.U., Kim, H., Kwon, Y., & Ko, H.J. (2015). Coadministration of *Hedera helix* L. Extract Enabled Mice to Overcome Insufficient Protection against Influenza A/PR/8 Virus Infection under Suboptimal Treatment with Oseltamivir. *PLoS One*, 10(6), e0131089. doi: 10.1371/journal.pone.0131089.
47. Ibrahim, N., & Moussa, A.Y. (2021). A comparative volatileomic characterization of Florence fennel from different locations: Antiviral Prospect. *Food & Function*. 12, 1498-1515. doi: 10.1039/DFO02897E.
48. Imran, M., Khan, S.A., Abida Alshammari, M.K, Alkhaldi, S.M., Alshammari, F.N., Kamal, M., Alam, O., Asdaq, S.M.B., Alzahrani, A.K., & Jomah, S. (2022). *Nigella sativa* L. and COVID-19: A Glance at The Anti-COVID-19 Chemical Constituents, Clinical Trials, Inventions, and Patent Literature. *Molecules*, 27(9),2750. doi: org/10.3390/molecules27092750.
49. Islam, Z., Caldeira, G.I., Caniça, M., Islam, N., Silva, O. (2024). Vitex Genus as a Source of Antimicrobial Agents. *Plants*, 13(3):401. doi: 10.3390/plants13030401.
50. James, A.D., Mary, Jo B-G., Judi du, C. & Peggy-Ann, K.D. (2003). CRC Handboook of medicinal spices. Boca Raton, London, New York, Washington: CRC Press.
51. Karagöz, A., Onay, E., Arda, N., & Kuru, A. (2003) Antiviral potency of mistletoe (*Viscum album* ssp. *album*) extracts against human parainfluenza virus type 2 in Vero cells. *Phytotherapy Research*, 17(5),560-562. doi: 10.1002/ptr.1163.
52. Karim, M., Lo, C. W., & Einav, S. (2023). Preparing for the next viral threat with broad-spectrum antivirals. *The Journal of Clinical Investigation*, 133 (11), e170236. doi: 10.1172/JCI170236.
53. Khodja, Y.K., Bachir-bey, M., Belmouhoub, M., Ladjouzi ,R., Dahmoune, F., & Khettal, B. (2023). The botanical study, phytochemical composition, and biological activities of *Laurus nobilis* L. leaves: A review. *International Journal of Secondary Metabolite*, 10(2), 269-296. doi: 10.21448/ijsm.1171836.
54. Kim, M., Nguyen, D.V., Heo, Y., Park, K.H., Paik, H.D., & Kim, Y.B. (2020). Antiviral Activity of *Fritillaria thunbergii* Extract against Human Influenza Virus H1N1 (PR8) In Vitro, In Ovo and In Vivo. *Journal of Microbiology and Biotechnology*, 30(2), 172-177. doi: 10.4014/jmb.1908.08001.

55. Kosari, M., Noureddini, M., Khamechi, S.P., Najafi, A., Ghaderi, A., Sehat, M., & Banafshe, H.R. (2021). The effect of propolis plus *Hyoscyamus niger* L. methanolic extract on clinical symptoms in patients with acute respiratory syndrome suspected to COVID-19: A clinical trial. *Phytotherapy Research*, 35(7), 4000-4006. doi: 10.1002/ptr.7116.
56. Kosari, M., Khorvash, F., Sayyah, M.K., Ansari Chaharsoughi, M., Najafi, A., Momen-Heravi, M., Karimian, M., Akbari, H., Noureddini, M., Salami, M., Ghaderi, A., Amini Mahabadi, J., Khamechi, S.P., Yeganeh, S., & Banafshe, H.R. (2024). The influence of propolis plus *Hyoscyamus niger* L. against COVID-19: A phase II, multicenter, placebo-controlled, randomized trial. *Phytotherapy Research*, 38(1), 400-410. doi: 10.1002/ptr.8032.
57. Kulkarni, S.A., Nagarajan, S.K., Ramesh, V., Palaniyandi, V., Selvam, S.P., & Madhavan, T. (2020). Computational evaluation of major components from plant essential oils as potent inhibitors of SARS-CoV-2 spike protein. *Journal of Molecular Structure*, 1221, 128823. doi: 10.1016/j.molstruc.2020.128823.
- 58.
59. Kulinović, M.F. (1898). Nešto o narodnom praznovjerju i liječenju u Muhamedovaca u Bosni Hercegovini. *Glasnik Zemaljskog Muzeja X/II-III:* 503-530.
60. Kurokawa, M., Shimizu, T., Watanabe, W., & Shiraki, K. (2010). Development of New Antiviral Agents from Natural Products. *Open Antimicrobial Agents Journal*, 2, 49-57. doi: 10.2174/18765181010020200049.
61. Kwon, D.H., Kwon, H.Y., Kim, H.J., Chang, E.J., Kim, M.B., Yoon, S.K., Song, E.Y., Yoon, D.Y., Lee, Y.H., Choi, I.S., & Choi, Y.K. (2005). Inhibition of hepatitis B virus by an aqueous extract of *Agrimonia eupatoria* L. *Phytotherapy Research*, 19(4), 355-358. doi: 10.1002/ptr.1689.
62. Lavoie, S., Côté, I., Pichette, A., Gauthier, C., Ouellet, M., Nagau-Lavoie, F., Mshvildadze, V., & Legault, J. (2017). Chemical composition and anti-herpes simplex virus type 1 (HSV-1) activity of extracts from *Cornus canadensis*. *BMC Complementary and Alternative Medicine*, 17, 123. doi: 10.1186/s12906-017-1618-2.
63. Mazurkova, N., Protsenko, M., Lobanova, I., Filippova, E., & Vysochina G. (2020). Antiviral activity of Siberian wild and cultivated plants. *BIO Web of Conferences* 24, 00051. doi.org/10.1051/bioconf/20202400051.
64. Medić, M. (1904a). Tri Ljekaruše. *Glasnik Zemaljskog Muzeja*. XVI/I:1-32.
65. Medić, M. (1904b). Tri Ljekaruše. (Nastavak i svršetak iz sveske I. str.1-32). *Glasnik Zemaljskog Muzeja*. XVI/II:195-223.
66. Mohamed, F.F., Anhlan, D., Schöfbänker, M., Schreiber, A., Classen, N., Hensel, A., Hempel, G., Scholz, W., Kühn, J., Hrincius, E.R., & Ludwig, S. (2022).

- Hypericum perforatum and Its Ingredients Hypericin and Pseudohypericin Demonstrate an Antiviral Activity against SARS-CoV-2. *Pharmaceuticals*, 15(5), 530. doi: 10.3390/ph15050530.
67. Monavari, S.H.R., Keyvani, H., & Bokharaei-Salim, F. (2012) Evaluation of in Vitro antiviral activity of Chelidonium majus L. against Herpes Simplex virus type-1. *African Journal of Microbiology Research*, 6(20), 4360-4364. doi:10.5897/AJMR11.1350.
68. Moradi, M.T., Karimi, A., Lorigooini, Z., Pourghneysari, B., Alidadi, S., & Hashemi, L. (2017). In vitro anti influenza virus activity, antioxidant potential and total phenolic content of twelve Iranian medicinal plants. *Marmara Pharmaceutical Journal*, 21(4), 843-851. doi: 10.12991/mpj.2017.10.
69. Muratović, E., & Parić, A. (2023). Plant ethnomedicine in Bosnia and Herzegovina, past and present. *Ethnobotany Research and Applications*, 26, 1–27. doi: 10.32859/era.26.61.1-27.
70. Musarra-Pizzo, M., Pennisi, R., Ben-Amor, I., Mandalari, G., & Sciortino, M.T. (2021). Antiviral Activity Exerted by Natural Products against Human Viruses. *Viruses*, 13(5), 828. doi: 10.3390/v13050828.
71. Musidlak, O., Warowicka, A., Broniarczyk, J., Adamczyk, D., Goździcka-Józefiak, A., & Nawrot, R. (2022). The Activity of Chelidonium majus L. Latex and Its Components on HPV Reveal Insights into the Antiviral Molecular Mechanism. *International Journal of Molecular Sciences*, 23(16), 9241. doi: 10.3390/ijms23169241.
72. Najar, B., Nardi, V., Stincarelli, M.A., Patrissi, S., Pistelli, L., & Giannecchini, S. (2022). Screening of the essential oil effects on human H1N1 influenza virus infection: an in vitro study in MDCK cells. *Natural Products Research*, 36(12), 3149-3152. doi: 10.1080/14786419.2021.1944137.
73. Najmi, A., Javed, S.A., Al Bratty, M., & Alhazmi, H.A. (2022). Modern approaches in the discovery and development of plant-based natural products and their analogues as potential therapeutic agents. *Molecules*, 27(2), 349. doi: 10.3390/molecules27020349.
74. Nikolaeva-Glomb, L., Mukova, L., Nikolova, N., Badjakov, I., Dincheva, I., Kondakova, V., Doumanova, L., & Galabov, A. (2014). In Vitro Antiviral Activity of a Series of Wild Berry Fruit Extracts against Representatives of Picorna-, Orthomyxo- and Paramyxoviridae. *Natural Product Communications*, 9, 51-54. doi: 10.1177/1934578X1400900116.
75. Oh, C., Price, J., Brindley, M.A., Widrllechner, M.P., Qu, L., McCoy, J.A., Murphy, P., Hauck, C., & Maury, W. (2011). Inhibition of HIV-1 infection by aqueous extracts of Prunella vulgaris L. *Virology Journal*, 8, 188. doi: 10.1186/1743-422X-8-188.

76. Okuda, T. (2005). Systematics and health effects of chemically distinct tannins in medicinal plants. *Phytochemistry*, 66, 2012–2031. doi: 10.1016/j.phytochem.2005.04.023.
77. Orhan, I., Ozçelik, B., Kartal, M., Özdeveci, B., Duman, H. (2007). HPLC Quantification of Vitexine-2"-O-rhamnoside and Hyperoside in Three *Crataegus* Species and Their Antimicrobial and Antiviral Activities. *Chromatographia*, 66, 153-157. doi: 10.1365/s10337-007-0283-x.
78. Orhan, İ.E., Özçelik, B., Kartal, M., & Kan, Y. (2012). Antimicrobial and antiviral effects of essential oils from selected Umbelliferae and Labiateae plants and individual essential oil components. *Turkish Journal of Biology*, 36, 239-246. doi: 10.3906/biy-0912-30.
- 79.
80. Pang, X., Zhao, J.Y., Wang, Y.J., Zheng, W., Zhang, J., Chen, X.J., Cen, S., Yu, L.Y., & Ma, B.P. (2020). Steroidal glycosides, homoisoflavanones and cinnamic acid derivatives from *Polygonatum odoratum* and their inhibitory effects against influenza A virus. *Fitoterapia*, 146, 104689. doi: 10.1016/j.fitote.2020.104689.
81. Parsania, M., Rezaee, M.B., Monavari, S.H., Jaimand, K., Mousavi-Jazayeri, S.M., Razazian, M., & Nadjarha, M.H. (2017). Antiviral screening of four plant extracts against acyclovir resistant herpes simplex virus type-1. *Pakistan Journal of Pharmaceutical Sciences*, 30(4(Suppl.)):1407-1411.
82. Perera, C., & Efferth, T. (2012). Antiviral Medicinal Herbs and Phytochemicals. *Journal of Pharmacognosy*, 3(1), 45-48. doi: 10.9735/0976-884X.3.1.45-48.
83. Pompei, R., Flore, O., Marccialis, M.A., Pani, A., & Loddo, B. (1979). Glycyrrhetic acid inhibits virus growth and inactivates virus particles. *Nature*, 281, 689–690. doi: 10.1038/281689a0.
84. Pujol, C.A., Damonte, E.B., Turjan, J., Yanbo, K.Z., & Capek, P. (2016). The antiviral potency of *Fagus sylvatica* 4OMe-glucuronoxylan sulfates. *Int J Biol Macromol*, 87, 195-200. doi: 10.1016/j.ijbiomac.2016.02.048.
85. Rajtar, B., Skalicka-Woźniak, K., Świątek, Ł., Stec, A., Boguszewska, A., & Polz-Dacewicz, M. (2017). Antiviral effect of compounds derived from *Angelica archangelica* L. on Herpes simplex virus-1 and Coxsackievirus B3 infections. *Food and Chemical Toxicology*, 109(Pt 2):1026-1031. doi: 10.1016/j.fct.2017.05.011.
86. Redžić, S. (2007). The ecological aspect of ethnobotany and ethnopharmacology of population in Bosnia and Herzegovina. *Collegium Antropologicum*, 31(3), 869-890.
87. Redžić, S. (2010). Wild medicinal plants and their usage in traditional human therapy (Southern Bosnia and Herzegovina, W. Balkan). *Journal of Medicinal Plants Research*, 4(11), 1003-1027. doi: 10.5897/JMPR09.254.

88. Rehman, S., Ijaz, B., Fatima, N., Muhammad, S.A., & Riazuddin, S. (2016). Therapeutic potential of *Taraxacum officinale* against HCV NS5B polymerase: In-vitro and In silico study. *Biomedicine & Pharmacotherapy*, 83, 881-891. doi: 10.1016/j.biopha.2016.08.002.
89. Salehi, B., Kumar, N.V.A., Şener, B., Sharifi-Rad, M., Kılıç, M., Mahady ,G.B., Vlaisavljevic, S., Iriti, M., Kobarfard, F., Setzer, W.N., Ayatollahi, S.A., Ata, A., & Sharifi-Rad, J. (2018). Medicinal Plants Used in the Treatment of Human Immunodeficiency Virus. *International Journal of Molecular Sciences*, 19(5):1459. doi: 10.3390/ijms19051459.
90. Sato, H., Goto, W., Yamamura, J., Kurokawa, M., Kageyama, S., Takahara, T., Watanabe, A., & Shiraki, K. (1996). Therapeutic basis of glycyrrhizin on chronic hepatitis B. *Antiviral Research*, 30, 171– 177. doi: 10.1016/0166-3542(96)00942-4.
91. Sekizawa, T., Yanagi, K., & Itoyama, Y. (2001). Glycyrrhizin increases survival of mice with herpes simplex encephalitis. *Acta Virologica*, 45, 51–54.
92. Serkedjieva, J., & Ivancheva, S. (1998). Antiherpes virus activity of extracts from the medicinal plant *Geranium sanguineum* L. *Journal of Ethnopharmacology*, 64, 59–68. doi: 10.1016/s0378-8741(98)00095-6.
93. Sharifi-Rad, J., Salehi, B., Schnitzler, P., Ayatollahi, S.A., Kobarfard, F., Fathi, M., Eisazadeh, M., & Sharifi-Rad, M. (2017). Susceptibility of herpes simplex virus type 1 to monoterpenes thymol, carvacrol, p-cymene and essential oils of *Sinapis arvensis* L., *Lallemandia royleana* Benth. and *Pulicaria vulgaris* Gaertn. *Cellular and Molecular Biology*, (Noisy-le-grand), 63(8), 42-47. doi: 10.14715/cmb/2017.63.8.10.
94. Ţtefănescu, B.E., Szabo, K., Mocan, A., & Crişan, G. (2019). Phenolic Compounds from Five Ericaceae Species Leaves and Their Related Bioavailability and Health Benefits. *Molecules*, 24(11), 2046. doi: 10.3390/molecules24112046.
95. Suganda, A.G., Amoros, M., Girre, L., & Fauconnier, B. (1983). Inhibitory effects of some crude and semi-purified extracts of indigenous French plants on the multiplication of human herpesvirus 1 and poliovirus 2 in cell culture. *Journal of Natural Products*, 46(5), 626-32. doi: 10.1021/np50029a006.
96. Sun, Y., Gong, X., Tan, J.Y., Kang, L., Li, D., Vikash Yang, J., & Du, G. (2016). In vitro Antiviral Activity of *Rubia cordifolia* Aerial Part Extract against Rotavirus. *Frontiers in Pharmacology*, 7, 308. doi: 10.3389/fphar.2016.00308.
97. Suručić, R., Radović Selgrad, J., Kundaković-Vasović, T., Lazović, B., Travari, M., Suručić, L., & Škrbić, R. (2022). In Silico and In Vitro Studies of *Alchemilla viridiflora* Rothm-Polyphenols' Potential for Inhibition of SARS-CoV-2 Internalization. *Molecules*, 27(16), 5174. doi: 10.3390/molecules27165174.

98. Sytar, O., Hemmerich, I., Zivcak, M., Rauh, C., & Brestic, M. (2018). Comparative analysis of bioactive phenolic compounds composition from 26 medicinal plants. *Saudi Journal of Biological Sciences*, 25(4), 631-641. doi: 10.1016/j.sjbs.2016.01.036.
99. Šarić-Kundalić, B., Ahmedbegović, A., Cilović, E., Ademović, Z., Kerleta-Tuzović, V., & Izić, B. (2015). Ethnobotanical study of traditionally used plants in human therapy of Trestenica and Tulovići, North-East Bosnia and Herzegovina. *Pharmacia*, 18(2):221-234.
100. Šarić-Kundalić, B., Dobeš, C., Klatte-Asselmeyer, V., Saukel, J. (2010a). Ethnobotanical study on medicinal use of wild and cultivated plants in middle, south and west Bosnia and Herzegovina. *Journal of Ethnopharmacology*, 131, 33-55. doi: 10.1016/j.jep.2010.05.061.
101. Šarić-Kundalić, B., Dobeš, C., Klatte-Asselmeyer, V., Saukel, J. (2011). Ethnobotanical survey of traditionally used plants in human therapy of east, north and north-east Bosnia and Herzegovina. *Journal of Ethnopharmacology*, 133, 1051-1076. doi: 10.1016/j.jep.2010.11.033.
102. Šarić-Kundalić, B., Fritz, E., Dobeš, C., & Saukel, J. (2010b). Traditional medicine in the pristine village of Prokoško lake on Vranica mountain, Bosnia and Herzegovina. *Scientia Pharmaceutica*, 78, 275-290. doi: 10.3797/scipharm.1003-06.
103. Šarić-Kundalić, B., Mazić, M., Djerić, S., & Kerleta-Tuzović, V. (2016). Ethnobotanical study on medicinal use of wild and cultivated plants on Konjuh Mountain, North-East Bosnia and Herzegovina. *Technics Technologies Education Management*, 11(3), 208- 221.
104. Takahara, T., Watanabe, A., & Shiraki, K. (1994). Effects of glycyrrhizin on hepatitis B surface antigen: a biochemical and morphological study. *Hepatology Research*, 21, 601–609. doi:10.1016/S0168-8278(94)80108-8.
105. Todorov, D., Pavlova, D., Hinkov, A., Shishkova, K., Dragolova, D., KapchinaToteva, V., & Shishkov, S. (2015) Effect of extracts derived from *Teucrium chamaedrys* L. on Herpes simplex virus type 2. *Dokladi na Bulgarskata Akademija na Naukite* 68, 1519–1526.
106. Tournier, J. N., & Kononchik, J. (2021). Virus eradication and synthetic biology: changes with SARS-CoV-2?. *Viruses*, 13(4), 569. doi: 10.3390/v13040569.
107. Trilla, A., Trilla, G., & Daer, C. (2008). The 1918 “Spanish Flu” in Spain. *Clinical Infectious Diseases*, 47(5), 668–673. doi:10.1086/590567.
108. Urda, L., Kreuter, M.H., Drewe, J., Boonen, G., Butterweck, V., & Klimkait, T. (2022). The *Petasites hybridus* CO₂ Extract (Ze 339) Blocks SARS-CoV-2 Replication In Vitro. *Viruses*, 14(1), 106. doi: 10.3390/v14010106.

- 109.Uncini Manganelli, R.E., Zaccaro, L., & Tomei, P.E. (2005). Antiviral activity in vitro of *Urtica dioica* L., *Parietaria diffusa* M. et K. and *Sambucus nigra* L. *Journal of Ethnopharmacology*, 98(3), 323-7. doi: 10.1016/j.jep.2005.01.021.
- 110.Vladimirov, M.S., Nikolić, V.D., Stanojević, Lj.P., Nikolić, Lj.B., & Tačić, A. (2019). Common birch (*Betula pendula* Roth.): Chemical composition and biological activity of isolates. *Advanced Technologies*, 8. doi: 10.5937/SavTeh1901065V.
- 111.Xiao, A, Syed, HA, Tsuchiya, A. *Eczema Herpeticum*. (2024). In: StatPearls [Internet]. Treasure Island (FL). StatPearls Publishing; Available from: <https://www.ncbi.nlm.nih.gov/books/NBK560781/>
- 112.Wang, L., Yang, R., Yuan, B., Liu, Y., & Liu, C. (2015). The antiviral and antimicrobial activities of licorice, a widely-used Chinese herb. *Acta Pharmaceutica Sinica B*, 5(4):310-315. 10.1016/j.apsb.2015.05.005.
- 113.Wang, H., Li, K., Ma, L., Wu, S., Hu, J., Yan, H., Jiang, J., & Li, Y. (2017). Berberine inhibits enterovirus 71 replication by downregulating the MEK/ERK signaling pathway and autophagy. *Virology Journal*, 14(1):2. doi: 10.1186/s12985-016-0674-4.
- 114.Wnorowska, S., Targowska-Duda, K., Kurzepa, J., Wnorowski, A., & Strzemski, M. (2022). Carlina oxide inhibits the interaction of SARS-CoV-2 S glycoprotein with angiotensin-converting enzyme 2. *Industrial Crops and Products*, 187, 115338. doi: 10.1016/j.indcrop.2022.115338.
- 115.World Health Assembly (2003). “Traditional Medicine: Report by the Secretariat.”. Fifty-sixth world health assembly, Provisional agenda item 14.10 (Geneva, Switzerland: World Health Organization). Available at: <https://apps.who.int/iris/handle/10665/78244>.
- 116.World Health Organization (2024). WHO COVID-19 dashboard. <https://data.who.int/dashboards/covid19/cases?n=o>
- 117.Zhakipbekov, K., Turgumbayeva, A., Issayeva, R., Kipchakbayeva, A., Kadyrbayeva, G., Tleubayeva, M., Akhayeva, T., Tastambek, K., Sainova, G., Serikbayeva, E., Tolenova, K., Makhatova, B., Anarbayeva, R., Shimirova, Z., & Tileuberdi, Y. (2023). Antimicrobial and Other Biomedical Properties of Extracts from *Plantago major*, Plantaginaceae. *Pharmaceuticals*, 16(8), 1092. doi: 10.3390/ph16081092.
- 118.Zhang, X., Jiang, Y., Qian, H., Qu, X., & Han, K. (2022). The association between Herpes simplex virus type 2 and asthma: A cross-sectional study from National Health and Nutrition Examination Survey 1999-2016. *Frontiers in Medicine*, 9, 943706. doi: 10.3389/fmed.2022.943706
- 119.Zhao, J. H., Wang, Y. W., Yang, J., Tong, Z. J., Wu, J. Z., Wang, Y. B., & Shi, Z. H. (2023). Natural products as potential lead compounds to develop new antiviral

- drugs over the past decade. European Journal of Medicinal Chemistry, 260:115726. doi: 10.1016/j.ejmech.2023.115726.
120. Zovko, I. (1890). Nekoliko narodnih lijekova. Glasnik Zemaljskog Muzeja, II/II:315-316.
121. Zumla, A., Hui, D. S., & Perlman, S. (2015). Middle East Respiratory Syndrome. Lancet, 386 (9997), 995–1007. doi:10.1016/S0140-6736(15)60454-8.